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A Comparative Study of Iron Oxide Nanoparticles Surface Modified Using Carboxylic Acids

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ABSTRACT

In the last decade nanotechnology has greatly developed in many research fields such as engineering, electronic, biological and many others. They can offer several possibilities to design tools, to create new techniques or improve the already existing ones, to discover innovative applications. Nano-science is one of the most important research and development frontiers in modern science. Nanotechnology is now widely used throughout the pharmaceutical industry, medicine, electronics, robotics, and tissue engineering. For biological and biomedical applications, magnetic iron oxide nanoparticles are the primary choice because of their biocompatibility, superparamagnetic behavior and chemical stability.

The purpose of this work is the design, development and surface modification of magnetic nanoparticles. Naked iron oxide nanoparticles have high chemical activity, toxicity and aggregate in the body fluid therefore providing surface coating for the stability of the magnetic nanoparticles. These protective shells not only stabilize the magnetic iron nanoparticles but also can be used for further functionalization. Here the iron oxide nanoparticles were prepared by co-precipitation method, then this nanoparticle is modified using acids- oleic acid and succinic acid and a comparative study is carried out. The TEM, FTIR and DSC characterization techniques were used to confirm the surface modification. After which, it was found the iron oxide nanoparticle with succinic acid gives a uniform coating of the three and can be used for further functionalization for various applications.

Keywords- Co-precipitation, Iron oxide nanoparticle, Oleic Acid, Citric Acid, Succinic Acid.

I. INTRODUCTION

Nanotechnology is defined as the creation and exploitation of materials with structural features in between those of atoms and bulk materials, of dimension in the nanometer range $(1 \text{ nm} = 10^{-9} \text{ m})$. A property of materials having nanometric dimensions significantly varies from those of atoms as well as those of bulk materials.

Nanotechnology covers fields from biology to material science, physics to chemistry, and can include development in a variety of specialities. Nanostructured materials have attracted considerable attention in recent years because they exhibit useful and unusual properties compared to conventional materials. They are of great interest in the creation of controlled micro and nanoarchitectures in an attempt to mimic the natural physical and biological environment that encourages tissue regeneration and growth. The hypothesis is that the nanoarchitectures can promote cell differentiation and functionality. By understanding how physical surface parameters influence cellular adhesion and differentiation we can more effectively design biomaterial surfaces for variety of tissue engineering applications. Further, nanostructured materials can be used as drug eluting interfaces for implantable devices, such as vascular stents, orthopedic implants, dental implants, etc. By precisely controlling the size of nanostructure, we can manipulate the release rates; thus releasing the drug at physiological levels.

1.1 Magnetic Nanoparticles

Magnetic nanoparticles of are a class nanoparticle, which can be manipulated using magnetic field. These particles commonly consist of magnetic elements such as iron, nickel and cobalt and their chemical compounds. The magnetic nanoparticles have been the focus of much research recently because they possess attractive properties which could see potential use in catalysis including nanomaterial-based catalysts,(1) biomedicine,(2)magnetic resonance $imaging_{(3)}$ magnetic particle imaging,(4)data storage,(5) environmental remediation,(6) nanofluids,(7) and optical filters,(8) defect sensor(9) and cation sensors.(10)

1.2 Iron Oxide Nanoparticles

Researchers are constantly working on new applications for nanoparticles in biological, chemical, and industrial areas. Iron oxides have a useful set of properties for biomedical imaging and catalytic applications. The two main forms are magnetite (Fe₃O₄) and its oxidized form maghemite (γ -Fe₂O₃). They have attracted extensive interest due to their superparamagnetic properties and their potential applications in many fields.

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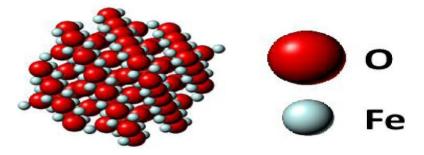


Figure 1.1: The crystal structure of iron oxide is a close-packed planes of oxygen anions (red) with iron cations (white) taking up some of the resulting octahedral or tetrahedral interstitial sites

Applications of iron oxide nanoparticles include terabit magnetic storage devices, catalysis, sensors, and high-sensitivity biomolecular magnetic resonance imaging (MRI) for medical diagnosis and therapeutics. These applications require coating of the nanoparticles by agents such as long-chain fatty acids, alkyl-substituted amines and diols.

Magnetic iron oxide (IO) nanoparticles with a long blood retention time, biodegradability and low toxicity have emerged as one of the primary nanomaterials for biomedical applications in vitro and in vivo. IO nanoparticles have a large surface area and can be surface modified to provide a large number of functional groups for cross-linking to tumor-targeting ligands such as monoclonal antibodies, peptides, or small molecules for diagnostic imaging or delivery of therapeutic agents. IO nanoparticles possess unique paramagnetic properties, which and generate significant susceptibility effects resulting in strong T_2 and T_2^* contrast, as well as T_1 effects at very low concentrations for magnetic resonance imaging (MRI), which is widely used for clinical oncology imaging. There are various methods currently used for the synthesis of nanomaterials. The following are a few common methods:

- a) Precipitation /wet chemical method
- b) Reduction of metal salt/ solution method
- c) Hydrothermal/solvothermal
- d) Hot thermolysis/colloidal synthesis

Here in my work I have used co-precipitation method for the synthesis of the nanoparticles.

This preparation method has a large effect on shape, size distribution, and surface chemistry of the particles. It also determines to a great extent the distribution and type of structural defects or impurities in the particles. All these factors affect magnetic behavior.

Co-precipitation

So far this is the most simple, cost effective and commonly employed method . As one convenient cheap method ,chemical co-precipitation has the potential to meet the increasing demand for the direct preparation of well dispersed nanoparticles. Chemical co-precipitation can produce fine ,high purity,stoichiometric particles of single and multicomponent metal oxides.

1.3 Surface Functionalization and It's Advantages

nanoparticles without any IO surface modification are not stable in aqueous media and readily aggregate and precipitate. For in vivo applications, the particles often form aggregates in blood and are sequestered by macrophages (Lee et al 2006). Therefore, the surface of IO nanoparticles should be coated with a variety of different moieties that can eliminate or minimize their aggregation under physiological conditions. Usually, two main approaches are used for coating magnetic IO nanoparticles, including in situ coatings with which the magnetic nanoparticles are coated during the synthesis process and post-synthesis coatings.(7).

The amphiphilic polymeric surfactants such as poloxamers, poloxamines and poly(ethylene glycol) (PEG) derivatives are usually used for coating the surface of IO nanoparticles, since they can minimize or eliminate opsonization of IO nanoparticles. Among them, PEG is the most used chemical material, which confers on IO nanoparticles several important properties such as high solubility and stability in aqueous solutions, biocompatibility, and prolonged blood circulation time.

Recently, a new class of superparamagnetic iron particles that have uniform sizes ranging from 5–30 was further functionalized through surface coating with amphiphilic polymers, which provide functional groups for conjugating tumor-targeting biomolecules such as peptides or antibodies(8). Hence surface modified magnetic nanoparticles are useful in biomedical applications. Here in my work I am coating the iron oxide nanoparticles using oleic acid and succinic acid.

II. EXPERIMENTAL PROCEDURE

2.1 General Description of the Work

First of all the iron oxide nanoparticles are synthesized using the co-precipitation method .As pure iron oxide nanoparticles are not stable in the aqueous media. IO nanoparticles without any surface modification does not have stability in aqueous media and readily aggregate and precipitate. For *in vivo* applications, the particles often form aggregates in blood and are sequestered by macrophages. Therefore, the surface of IO nanoparticles should be coated with a variety of different moieties that can eliminate or minimize their aggregation under physiological conditions. Usually, two main approaches are used for coating magnetic IO nanoparticles, including *in situ* coatings with which the magnetic nanoparticles are coated during the synthesis process and post-synthesis coatings. Here the first one is done and the procedure is as explained below.

2.1.1 Synthesis of Pure Iron Oxide Nanoparticles Materials and Reagents

Ferric Chloride Hexahydrate (FeCl₃.6H₂O), (Nice Labortary, Mumbai), Iron (II) sulphate Heptahydrate GR (Ferrous sulphate) (Merck, Mumbai), Liquor ammonia (Qualigens Fine Chemicals, Mumbai) *Methodology*

 Fe_3O_4 nanoparticles were synthesized by coprecipitation method consisting of FeCl₃.6H₂O (2.36g) and FeSO₄.7H₂O (0.86g) at the molar ratio 2:1 and they were dissolved in deionized water. After stirring for 10 minutes, NH₄OH (3.2 M) was then added to the solution which was kept for three hours of continuous stirring. The Fe₃O₄ nanoparticles were separated centrifugally at 15000rpm for 10 minutes and washed three times with water.

The above synthesized nanoparticle was considered for the further surface modifications.

III. SURFACE MODIFIED IRON OXIDE NANAOPARTICLES

3.1 Synthesis of Iron Oxide Nanoparticles Coated with Oleic Acid

Materials and Reagents

Ferric Chloride Hexahydrate (FeCl₃.6H₂O), (Nice Labortary, Mumbai, Iron (II) sulphate Heptahydrate GR (Ferrous sulphate) (Merck, Mumbai), Liquor ammonia (Qualigens Fine Chemicals, Mumbai), Oleic Acid (Merck, Germany).

Methodology

FeCl₃.6H₂O (3g) and FeSO₄.7H₂O (1.5g) were dissolved in almost 5ml of distilled water. Then this iron salt solution was kept for stirring for 5 minutes at 70° C in a nitrogen purged atmosphere. Then after 5 minutes, 5ml of liquid ammonia solution was added into the flask which gives a black precipitate which is followed by the addition of 450µl of oleic acid. This oleic acid dispersed solution was kept for continuous stirring for three hours in nitrogen atmosphere. After, three hours of continuous stirring we observe a black solution with precipitates. This solution was then allowed to cool at room temperature for almost 20 minutes which was followed by the centrifugation inorder to remove the excess salts. . The resultant was then centrifuged at 8000rpm at $4C^0$ for 20 minutes and later centrifuged at 15000rpm at 4 degree for 50 minutes. The resultant sample after centrifugation was kept for lyophilization. The nanoparticles thus obtained were characterized using TEM and FTIR in order to know the particle size and to confirm the chemical adsorption of -COOH groups onto nanoparticle surface.

3.2 Synthesis of Iron Oxide Nanoparticles Coated with Citric Acid

Materials and Reagents

Ferric Chloride Hexahydrate (FeCl₃.6H₂O), (Nice Labortary, Mumbai), Iron (II) sulphate Heptahydrate GR (Ferrous sulphate) (Merck, Mumbai), Liquor ammonia (Qualigens Fine Chemicals, Mumbai), Citric Acid (Sigma Aldrich, USA).

Methodology

FeCl₃.6H₂O (3g) and FeSO₄.7H₂O (1.5g) were dissolved in almost 5ml of distilled water. Then this iron solution was kept for stirring for 5minutes at 70° C in a nitrogen purged atmosphere. Then after 5 minutes, 5ml of liquor ammonia solution was added into the flask which gives a black precipitate which is followed by the addition of 5g of citric acid dissolved in 10 ml of distilled water. This citric acid dispersed solution was kept for continuous stirring for three hours in nitrogen atmosphere. After, three hours of continuous stirring, this solution was then allowed to cool at room temperature for almost 20 minutes which was followed by the dialysis procedure inorder to remove the excess salts. After the dialysis procedure the resultant solution was kept for lyophilization were the solution will be changed into powder form by extracting the water inorder to remain stable and to make it easier to store at room temperature. The obtained nanoparticles are characterized using TEM and FTIR to know the particle size and to confirm the chemical adsorption of -COOH groups onto the nanoparticle surface.

3.3 Synthesis of Iron Oxide Nanoparticles Coated with Succinic Acid

Materials and Reagents

Ferric Chloride Hexahydrate (FeCl₃.6H₂O), (Nice Labortary, Mumbai), Iron (II) sulphate Heptahydrate GR (Ferrous sulphate) (Merck, Mumbai), Liquor ammonia (Qualigens Fine Chemicals, Mumbai), Succinic Acid (Sigma Aldrich, USA).

Methodology

FeCl₃.6H₂O (3g) and FeSO₄.7H₂O (1.5g) were dissolved in almost 5ml of distilled water. Then this iron solution was kept for stirring for 5 minutes at 70° C in a nitrogen purged atmosphere. Then after 5 minutes, 5ml of liquor ammonia solution was added into the flask which gives a black precipitate which is followed by the addition of 5g of succinic acid dissolved in 10 ml of distilled water. This succinic acid dispersed solution was kept for continuous stirring for three hours in nitrogen atmosphere. After, three hours of continuous stirring, this solution was then allowed to cool at room temperature for almost 20 minutes which was followed by the dialysis procedure inorder to remove the excess salts. After the dialysis procedure the resultant solution was kept for lyophilization were the solution will be changed into powder form by extracting the water inorder to remain stable and to make it easier to store at room temperature. The obtained nanoparticles are characterized using TEM and FTIR to know the particle size and to confirm the chemical adsorption of -COOH groups onto the nanoparticle surface.

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IV. RESULTS AND DISCUSSIONS

4.1 Tem Analysis

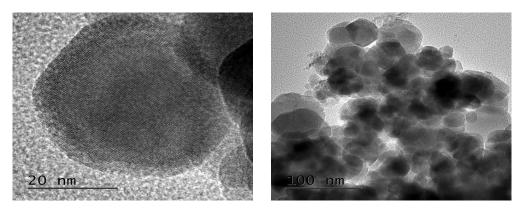


Figure 4.1.1: Pure Iron oxide nanoparticles

Inferences:

The TEM image of the iron oxide nanoparticle confirms the morphology and size of the nanoparticle.

The TEM image shows that the particles are aggregated because the metal oxide nanoparticles tend to react more with each other.

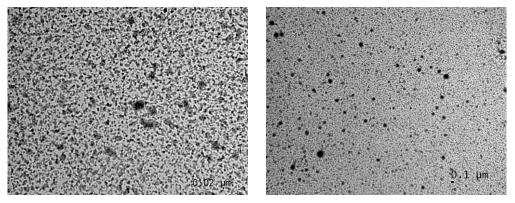


Figure 4.1.2: Iron oxide nanoparticles coated with oleic acid

Inferences:

Figures above shows TEM images and size distribution of iron oxide nanoparticles coated with oleic acid. It can be seen that plain iron oxide nanoparticles were polydisperse and seriously aggregated. After surface modification by oleic acid the particles maintained their

original spherical shape with a good monodispersity. The particle size is very uniform with the average size of about 100 nm- 200nm. From the magnified image due to the presence of excess oleic acid the aggregation of individual particles with an incomplete coating by oleic acid molecules can be observed.

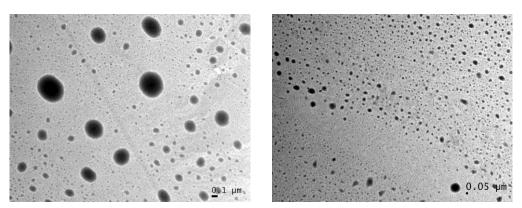


Figure 4.1.3: Iron oxide nanoparticles coated with succinic acid

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Inferences:

After surface modification by citric acid the particles maintained their original spherical shape with a good monodispersity. The particle size is very uniform with the average size of about 50nm - 200nm. It can be observed that the excess of citric acid is present hence this surface coating was also neglected.

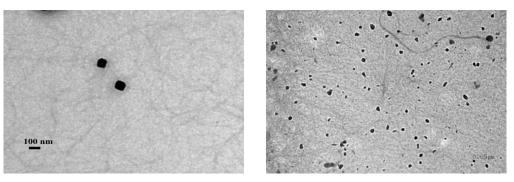


Figure 4.1.4: Iron oxide nanoparticles coated with succinic acid

Inferences:

Figures above shows the TEM images and size distribution of iron oxide nanoparticles coated with succinic acid. It can be seen that plain iron oxide nanoparticles were polydisperse and seriously aggregated. After surface modification by succinic acid the particles **4.2 Ftir Analysis**

maintained their original spherical shape with a good monodispersity. The particle size is very uniform with the average size of about 50nm- 100nm. It can be observed that the excess of succinic acid is present but that was washed off using dialysis procedure.

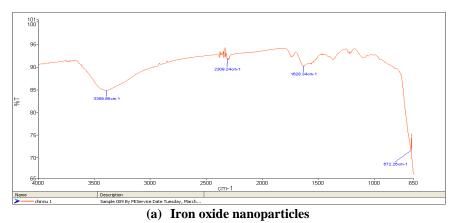


Figure 4.2.1: Pure iron oxide nanoparticles

Inferences:

Figure above shows the FTIR result of pure iron oxide nanoparticles. Here the analysis indicated

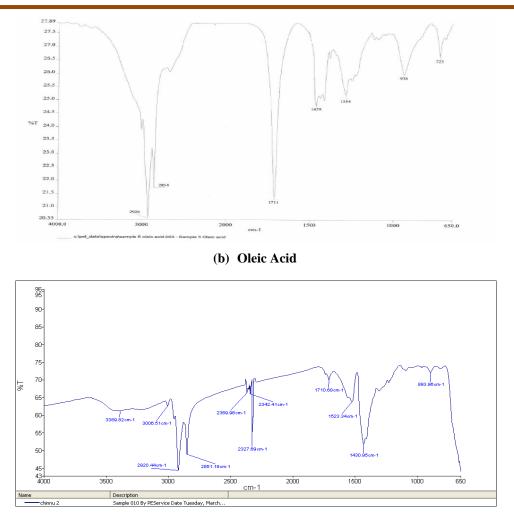
absorption peaks at 672.26 cm^{-1} , 1629.34 cm^{-1} , 2309.24 cm^{-1} and 3389.86 cm^{-1} corresponding to the Fe–O vibration related to the magnetite phase.



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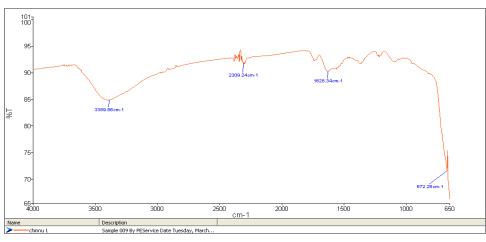
(c) Iron oxide nanoparticles coated with oleic acid

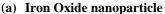
Figure 4.2.2: Comparison of FTIR Analysis of (a) Iron oxide nanoparticle, (b) Oleic Acid and (c) Iron oxide nanoparticle coated with Oleic Acid

Inferences:

The two absorption peaks at 2920.44 cm⁻¹ and 2,851.19 cm⁻¹ were attributed to the asymmetric CH₂ stretching and the symmetric CH₂ stretching,

respectively. The intense peaks at 2327.69 cm^{-1} and 1430.95 cm^{-1} was due to the overlapping of the absorption bands of the carboxyl groups and the double bonds of OA.





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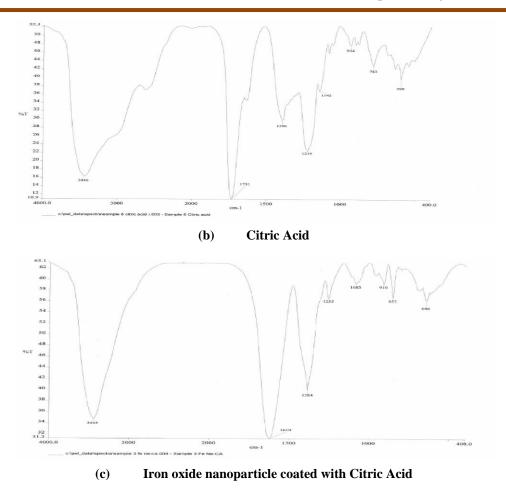
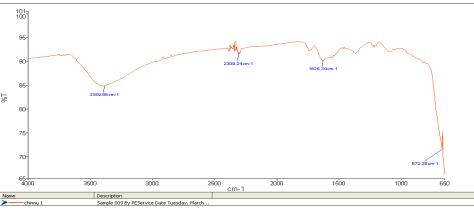


Figure 4.2.3: Comparison of FTIR Analysis of (a) Iron oxide nanoparticle, (b) Citric Acid and (c) Iron oxide nanoparticle coated with Citric Acid

Inferences:

The absorption bands for pure citric acid are resolved but those of iron oxide nanoparticle coated with citric acid are broad and few. The intense band at 3446 cm⁻¹shows the presence of non dissociated OH groups of citric acid .The peak around 1731 cm⁻¹ is due to the CH₂ stretching ,peak at 1396 cm⁻¹may be due to the symmetric stretching of OH from –COOH group. The peak at 1700cm⁻¹ of citric acid shows the C=O vibration from the

COOH group of citric acid. This peak shifts to an intense band at about 1255cm^{-1} showing the binding of citric acid onto the magnetite nanoparticles. The next band at 1142cm^{-1} can be due to the asymmetric stretching of CO from COOH group. The strong FTIR band observed at around 598cm^{-1} can be due to the Fe-O stretching vibration of Fe₃O₄. Therefore, we can say that the citric acid binds to the magnetite surface by carboxylate.



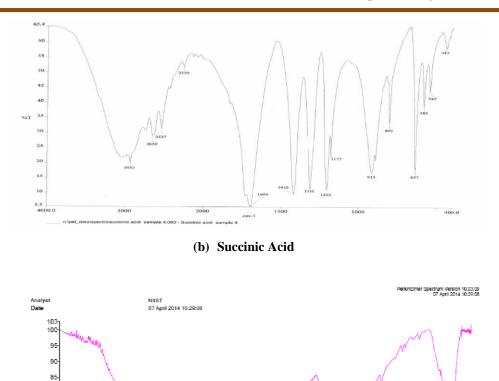




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Inferences:

Figure above shows the FTIR spectrum for the succinic acid coated nanoparticles. The spectrum confirms the presence of Fe-O stretches peaks around 580 and 620 cm⁻¹. The peaks at 3650 cm⁻¹ shows the presence of non dissociated OH groups of succinic acid. Compared

15.53

3500

3000

2500

(c) Iron oxide nanoparticles coated with succinic acid

Figure 4.2.4 Comparison of FTIR Analysis of (a) Iron oxide nanoparticle, (b) Succinic Acid and (c) Iron oxide nanoparticle coated with Succinic Acid

2000

cm-1

1500

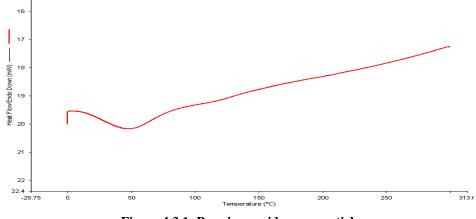
80-75-70-65-60-59-4000

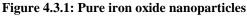
to pure succinic acid, there is a shift in the peak at about 1646cm^{-1} for iron oxide nanoparticles coated with succinic acid displaying the binding of the succinic acid onto the surface of Fe₃O₄ nanoparticles by chemiabsorption of carboxylate ions.

1000

500400

4.3 Differential Scanning Calorimetry







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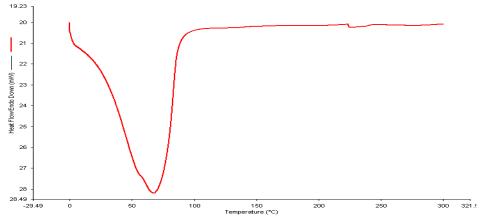
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DSC curve gives us the temperature at the midpoint of the

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Inferences:

The DSC curve of pure iron oxide nanoparticle is as shown above. It shows an exothermic curve. The



curve as 47.55° .

Figure 4.3.2: Iron oxide nanoparticles coated with succinic acid

Inferences:

Figure shows the DSC curve of iron oxide nanoparticle coated with succinic acid. The curve indicates an exothermic one. The temperature at the midpoint of the peak of iron oxide nanoparticle coated with succinic acid is 67.77° . So comparing with the DSC curve of pure iron oxide nanoparticle (figure 4.3.1), the curve gets shifted which shows that succinic acid binds to the magnetite surface by carboxylate.

V. **CONCLUSION**

The work investigated the synthesis of superparamagnetic iron oxide nanoparticles prepared by the simple and cost effective method, co-precipitation method. As the naked iron oxide nanoparticles cannot be used for the in vitro applications as they are toxic, agglomerates in the body fluid. To overcome this disadvantage the iron oxide nanoparticles were surface modified. The nanoparticles were surface modified using three types of carboxylic acids -oleic acid, citric acid and succinic acid. A comparative study was carried out among them using TEM, FTIR and DSC characterization techniques. Out of the three, iron oxide nanoparticle coated with succinic acid gives a uniform coating among them which was confirmed using TEM analysis and we came to a conclusion that we can use these nanoparticles for further functionalization which can be used for various applications.

REFERENCES

[1] A.-H. Lu, W. Schmidt, N. Matoussevitch, H. Bönnemann, B. Spliethoff, B. Tesche, E. Bill, W. Kiefer, F. Schüth (August 2004). "Nanoengineering of a Magnetically Separable Hydrogenation Catalyst". Angewandte Chemie International Edition 43 (33): 4303-4306.

[2] A.K. Gupta, M. Gupta (June 2005). "Synthesis and surface engineering of iron oxide nanoparticles for biomedical applications". Biomaterials 26 (18): 3995-4021.

[3] S. Mornet, S. Vasseur, F. Grasset, P. Verveka, G. Goglio, A. Demourgues, J. Portier, E. Pollert, E. Duguet (2006). Prog. Solid State Chem. 34: 237.

[4] B. Gleich, J. Weizenecker (2005). "Tomographic imaging using the nonlinear response of magnetic particles". Nature 435 (7046): 1214-1217. T. Hyeon (2003). Chem. Commun.: 927.

[5] D. W. Elliott, W.-X. Zhang (2001). Environ. Sci. Technol. 35: 4922.

[6] J. Philip, Shima.P.D. B. Raj (2006). "Nanofluid with tunable thermal properties". Applied Physics Letters 92: 043108.

[7] J.Philip, T.J.Kumar, P.Kalyanasundaram, B.Raj (2003). "Tunable Optical Filter". Measurement Science & Technology 14: 1289–1294.

[8] V. Mahendran and J.Philip "Nanofluid based opticalsensor for rapid visual inspection of defects in materials"Appl. ferromagnetic Phys. Lett. 100. 073104(2012);

[9] J.Philip, V. Mahendran, and Leona J. Felicia "A Simple, In-Expensive and UltrasensitiveMagnetic Nanofluid Based Sensor for Detection of Cations, Ethanol and Ammonia "J. Nanofluids 2, 112-119 (2013)

[10] Teja, Amyn S.; Koh, Pei-Yoong (2009). "Synthesis, properties, and applications of magnetic iron oxide nanoparticles". Progress in Crystal Growth and Characterization of Materials 55: 22.

[11] C. V. Thach, N. H. Hai and N. Chau," Size Controlled Magnetite Nanoparticles and Their Drug Loading Ability "Journal of the Korean Physical Society, (2008), 1332-1335,vol 5

[12] Sophie Laurent1, Morteza Mahmoudi, "Superparamagnetic iron oxide nanoparticles: promises

for diagnosis and treatment of cancer"Int J Mol Epidemiol Genet 2011;2(4):367-390

[13] Mahnaz Mahdavi , Mansor Bin Ahmad , Md Jelas Haron , Farideh Namvar, Behzad Nadi , Mohamad Zaki Ab Rahman and Jamileh Amin ," Synthesis, Surface Modification and Characterisation of Biocompatible Magnetic Iron Oxide Nanoparticles for Biomedical Applications" *Molecules* 2013, *18*, 7533-7548

[14] H. Asnaashari Eivari, A. Rahdar, H.Arabi et al (2012),"Preparation of supermagnatic iron oxide nanoparticles and investigating their magnetic properties" Vol 5 131-135

[15] K. Kim, Y. Zhang, W. Voit, K.V. Rao, M. Muhammed(2008)"A simple way to synthesize superparamagnetic iron oxide nanopartilces prepared by co-precipitation technique"Vol 6 543-612

[16] M. Răcuciu, D.E. Creangă, A. Airinei, D. Chicea, V. Bădescu, "Synthesis and properties of magnetic nanoparticles coated with biocompatible compounds"*Materials Science-Poland, Vol. 28, No. 3, 2010*

[17] Alona Gabrene, Janina Setina, Inna Juhnevica and Gundars Mezinskis, "Stabilization of Magnetite Nanoparticles by Encapsulation into the Silica Matrix"J Chem. Eng. 8 (2014) 42-46

[18] Lind M.J., M.J. (2008). "Principles of cytotoxic chemotherapy". *Medicine* 36 (1): 19–23.

[19] Nastoupil, LJ; Rose, AC; Flowers, CR (2012). "Diffuse large B-cell lymphoma: current treatment approaches". *Oncology (Williston Park, N.Y.)* 26 (5): 488–95.

[20] Freedman, A (2012). "Follicular lymphoma: 2012 update on diagnosis and management". *American journal of hematology* 87 (10): 988–95.

[21] Rampling, R; James, A; Papanastassiou, V (2004). "The present and future management of malignant brain tumours: surgery, radiotherapy, chemotherapy". *Journal of neurology, neurosurgery, and psychiatry*. 75 Suppl 2 (Suppl 2): ii24–30..

[22] Madan, V; Lear, JT; Szeimies, RM (2010). "Nonmelanoma skin cancer". *Lancet* 375 (9715): 673–85.

[23] CK Bomford, IH Kunkler, J Walter. Walter and Miller's Textbook of Radiation therapy (6th Ed), p311

[24] Jongnam Park, Jin Joo, Soon Gu Kwon, Youngjin Jang, and Taeghwan Hyeon (2007) J.Chem. "A Mechanism for the adsorption of carboxylic acids onto the surface of magnetic nanoparticles", Int. Ed. 2007, 46, 4630 – 4660.

[25] S. Ayyappan, G. Gnanaprakash, G. Panneerselvam, M.P. Antony, and John Philip, "Intracellular trafficking of superparamagnetic iron oxide nanoparticles conjugated with TAT peptide: 3-dimensional electron tomography analysis" *J. Phys. Chem. C* 2008, *112*, 18376–18383

[26] Xun Wang, Jing Zhuang, Qing Peng and Yadong Li ," Design and fabrication of magnetic nanoparticles for targeted drug delivery and imaging"Nature, Vol 437, 121-124, 1 September 2005. https://doi.org/10.31033/ijrasb.8.1.13

[27] María Elisa de Sousa, Marcela Beatriz Fernandez van Raap, Patricia Claudia Rivas, Pedro Mendoza Zélis,Pablo Girardin, Gustavo A. Pasquevich, Jose L. Alessandrini, Diego Muraca, and Francisco H Sánchez "Hybrid magnetic nanostructures (MNS) for magnetic resonance imaging applications" J. Phys. Chem. C,Feb 2013

[28] Maria E. N. P. Ribeiro ,Nágila M. P. S. Ricardo , Tamara Gonçalves , Luigi Carbone , Telma L. G. Lemos ,Otília D. L. Pessoa and Pierre B. A. Fechine , "multimodal spion-creka peptide based agents for molecular imaging of microthrombus in a rat myocardial ischemia-reperfusion model"*int. j. mol. sci.* 2013, *14*, 18269-18283

[29] L. Li , K.Y. Mak, C.W. Leung , K.Y. Chan , W.K. Chan , W. Zhong , P.W.T. Pong , "Engineering of cell penetrating peptides over iron oxide nanopartilces" Microelectronic Engineering 110 (2013) 329–334

[30] K.R.Vinodh, D.Sandhya, D.Banji and T. Rohit Reddy ."Application of iron oxide nanopartilces in tumor targeting and imaging "International Journal of Pharmaceutical Sciences and Nanotechnology, Vol 4, Issue 4 January-March 2012

[31] Sutima Chatrabhuti, Suwabun Chirachanchai, "Surface Functionalization of different polymers and other surfactants on the iron oxide nanopartilces" Carbohydrate Polymers 97 (2013) 441–450

[32] L. Borlido , A.M. Azevedo , A.C.A. Roque , M.R. Aires-Barros , "Nanoparticle imaging for MRI and NMR researchers" Biotechnology Advances 31 (2013) 1374–1385

[33] Veerle Kersemans and Bart Cornelissen, "*Effect of ligand density,target density and size on the nanoparticle*" *Pharmaceuticals* 2010, *3*, 600-620

[34] Maarten Bloemen ,Ward Brullot ,Tai Thien Luong , Nick Geukens , Ann Gils ,Thierry Verbiest, "Synthesis and applications of iron oxide nanoparticles in biomedical fields"J Nanopart Res (2012) 14:1100N.

[35] Nitin Æ L. E. W. LaConte Æ O. Zurkiya Æ X. Hu, G. Bao, "Effect of the surface ligands and its different applications in different fields" J Biol Inorg Chem (2004) 9: 706–712